

Review of the Glutathione Depletion—Methylation Cycle Block (GD-MCB) Hypothesis for CFS [1]

- 1. The person inherits a genetic predisposition (polymorphisms in several of certain genes) toward developing CFS. (This genetic factor is more important for the sporadic cases than for the cluster cases of CFS.)
- 2. The person then experiences some combination of a variety of possible stressors (physical, chemical, biological, psychological/emotional) that place demands on glutathione.
- 3. Glutathione levels drop, producing oxidative stress, removing protection from B12, allowing toxins to accumulate, and shifting the immune response to Th2.
- 4. Toxins react with B12, lowering the rate of formation of methylcobalamin.
- Lack of sufficient methylcobalamin inhibits methionine synthase, placing a partial block in the methylation cycle.
- 5. Sulfur metabolites drain through the transsulfuration pathway excessively, pass through sulfoxidation, and are excreted.
- 6. A vicious circle is established between the methylation cycle block and glutathione depletion, and the disorder becomes chronic.

Depletion of glutathione by *Borrelia burgdorferi*

- 1. Bb requires cysteine for its metabolism [2].
- 2. Cysteine diffuses passively into Bb from its host, i.e. there is no active transporter protein [2].
- 3. Bb uses cysteine in the synthesis of several of its essential enzymes: Osp A, Osp B, CoASH, a hemolysin, and others [2,3].
- 4. Bb does not use glutathione for its redox control. Instead, it uses reduced Coenzyme A (CoASH) [4].
- 5. Cysteine is the rate-limiting amino acid for the synthesis of glutathione in humans, so that depletion of cysteine will produce depletion of glutathione [5].
- 6. Bb lowers the cysteine and glutathione levels in its human host, and inhibits the activity of glutathione peroxidase [6].
- 7. Low glutathione and low activity of glutathione peroxidase allow a rise in hydrogen peroxide concentration and oxidative stress [7].
- 8. Elevation of hydrogen peroxide causes Bb to assume its cyst form [8], in which it is less vulnerable to antibiotics [9].

New hypothesis for a link between Lyme disease and chronic fatigue syndrome

- 1. *Borrelia burgdorferi* (Bb) deplete glutathione in the host.
- 2. For a person who is genetically susceptible to developing CFS, this provides a link to the GD-MCB hypothesis for CFS and is one of the possible routes into this disorder.

- 3. If Bb and its biotoxin were not eliminated, Lyme disease and CFS would coexist in the host, and this would constitute “chronic Lyme disease.”
- 4. If Bb and its biotoxin [10] were eliminated, but the methylation cycle block continued, the person would continue to be ill with CFS. This would constitute “post-Lyme disease syndrome,” which would be analogous to the other post-infective fatigue syndromes [11].
- 5. If Bb and its biotoxin were eliminated, and the methylation cycle block was lifted, I believe it is likely that the person would become well.

In addition,

- 6. Perhaps the *Borrelia burgdorferi* toxin is one of the toxins that will react with vitamin B12. Mold toxins have been implicated in such reactions, but no data were cited [12,13].

References

1. Van Konynenburg, R.A., “Glutathione Depletion—Methylation Cycle Block, A Hypothesis for the Pathogenesis of Chronic Fatigue Syndrome,” poster paper, 8th Intl. IACFS Conf. on CFS, Fibromyalgia, and Other Related Illnesses, Fort Lauderdale, FL, January 10-14, 2007
<http://phoenix-cfs.org/GSHMethylationVanKonynenburg.htm>
2. Sambri, V., and Cevenini, R., Incorporation of cysteine by *Borrelia burgdorferi* and *Borrelia hersii*, *Can. J. Microbiol.* 38: 1016-1021 (1992).
3. Williams, L.R., and Austin, F.E., Hemolytic activity of *Borrelia burgdorferi*, *Infection and Immunity* 60(8): 3224-3230 (1992).
4. Boylan, J.A., Hummel, C.S., Benoit, S., Garcia-Lara, J., Treglown-Downey, J., Crane, E.J., III, and Gherardini, F.C., *Borrelia burgdorferi* bb0728 encodes a coenzyme A disulphide reductase whose function suggests a role in intracellular redox and the oxidative stress response, *Molecular Microbiol.* 59(2), 475-486 (2006).
5. Griffith, O.W., Biologic and pharmacologic regulation of mammalian glutathione synthesis, *Free Radic Biol Med.* 1999 Nov;27(9-10):922-35.
6. Pancewicz, S.A., Skrzydlewska, E., Hermanowska-Szpakowicz, T., Zajkowska, J., and Kondrusik, M., Role of reactive oxygen species (ROS) in patients with erythema migrans, an early manifestation of Lyme borreliosis, *Med. Sci. Monit.* 7(6), 1230-1235
7. Levine, S.A., and Kidd, P.M., *Antioxidant Adaptation, Its Role in Free Radical Pathology*, Allergy Research Group, San Leandro, CA (1985).
8. Murgia, R., and Cinco, M., Induction of cystic forms by different stress conditions in *Borrelia burgdorferi*, *APMIS* 112, 57-62 (2004).
9. Kersten, A., Poitschek, C., Rauch, S., and Aberer, E., Effects of penicillin, ceftriaxone and doxycycline on morphology of *Borrelia burgdorferi*, *Antimicrob. Agents Chemother.* 39(5), 1127-1133 (1995).
10. Shoemaker, R., Schaller, J., and Schmidt, P., Mold Warriors, Gateway Press, Baltimore (2005).
11. Hickie, I. Davenport, T., Wakefield, D, Vollmer-Conna, U., Cameron, B., Vernon, S.D., Reeves, W.C., Lloyd, A., Dubbo Infection Outcomes Study Group, Post-infective

and chronic fatigue syndromes precipitated by viral and non-viral pathogens: prospective cohort study, *BMJ*. 2006 Sep 16;333(7568):575. Epub 2006 Sep 1.

12. Anyanwu, E.C., Morad, M., and Campbell, A.W., Metabolism of mycotoxins, intracellular functions of vitamin B12, and neurological manifestations in patients with chronic toxigenic mold exposures. A review, *ScientificWorldJournal* 4, 736-745 (2004).

13. Anyanwu, E.C., and Kanu, I., Biochemical impedance on intracellular functions of vitamin B12 in chronic toxigenic mold exposures, *ScientificWorldJournal* 7:1649-57 (2007).